USER MANUAL

CULTURE MYOGRAPH SYSTEM

MODEL 204CM

User Manual

Version 1.7

CULTURE MYOGRAPH SYSTEM
MODEL 204CM

TRADEMARKS

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This document was, as far as possible, accurate at the time of printing.
Changes may have been made to the software and hardware described since then.
New information may be supplied separately.

This documentation is provided with the DMT Culture Myograph System – Model 204CM

Document Number: 204CM 001A

Every attempt is made to ensure accurate information, misprints, construction- and specification changes can occur.
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CONTENTS

SAFETY

The Culture Myograph System has been designed for use only in teaching and research applications. It is not intended for clinical or critical life-care use and should never be used for these purposes: nor for the prevention, diagnosis, curing, treatment, or alleviation of disease, injury or handicap.

- Do not open the unit: the internal electronics pose a risk of electric shock.
- Do not use this apparatus near water.
- To reduce the risk of fire or electric shock, do not expose this apparatus to rain or moisture. Objects filled with liquids should not be placed on the apparatus.
- Do not block any ventilation openings. Install in accordance with the manufacturer's instructions.
- Do not install near any heat sources such as radiators, heat registers, stoves, or other apparatus that produce heat.
- Only use attachments and accessories specified by the manufacturer.
- Unplug this apparatus during lightning storms or when unused for long periods of time.
- This apparatus must be earthed.
- Use a three-wire grounding-type cord similar to the one supplied with the product.
- Do not defeat the safety purpose of the polarized or grounding-type plug. A polarized plug has two flat blades, one being wider than the other. A grounding type plug has two blades and a third (round) grounding pin. The wide blade or the third prong is provided for your safety. If the provided plug does not fit into your outlet, consult an electrician for replacement of the obsolete outlet.
- Be advised that different operating voltages require the use of different types of line cord and attachment plugs. Check the voltage in your area and use the correct type. See the table below:

Voltage	Line plug according to standard
110–125 V	UL817 and CSA C22.2 No. 42.
220–230 V	CEE 7 page VII, SR section 107-2-D1/IEC 83, page C4.
240 V	BS 1363 of 1984. Specification for 13A fused plugs and switched and unswitched socket outlets.

Protect the power cord from being walked on or pinched: particularly at power plugs and the point where they connect to the apparatus.

Refer all servicing to qualified service personnel. Servicing is required when the apparatus has been damaged in any way; such as, the power-supply cord or plug is damaged, liquid has been spilled onto or objects have fallen into the apparatus, the apparatus has been exposed to rain or moisture, does not operate normally, or has been dropped.

EMC/EMI

This equipment has been tested and found to comply with the limits for a Class B Digital device, pursuant to part 15 of the FCC rules. These limits are designed to provide reasonable protection against harmful interference in residential installations. This equipment generates, uses and can radiate radio frequency energy and, if not installed and used in accordance with the instructions, may cause harmful interference to radio communications. However, there is no guarantee that interference will not occur in a particular installation. If this equipment does cause harmful interference to radio or television reception (which can be determined by monitoring the interference while turning the equipment off and on), the user is encouraged to correct the interference by one or more of the following measures:

- Reorient or relocate the receiving antenna.
- Increase the separation between the equipment and receiver.
- Connect the equipment into an outlet on a circuit different to that to which the receiver is connected to.
- Consult the dealer or an experienced radio/TV technician for help.

APPROVALS

Complies with the EMC standards:
EMC 89/336/EEC: EN 50 081-1 and EN 50 082-1
FCC part 15, Class B
CISPR 22, Class B

Certified with the safety standards:
EN 60 065 (IEC 60065)

Complies with the safety standards:
UL6500
CSA E65

CERTIFICATE OF CONFORMITY

DMT A/S, Skejbyparken 152, 8200 Aarhus N., Denmark,
hereby declares its responsibility that the following product:

Culture Myograph System– Model 204CM

is covered by this certificate and marked with CE-label and conforms with the following standards:

EN 60 065 (IEC 65)	Safety requirements for mains operated Electronic and related apparatus for household and similar general use.
EN 50 081-1	Electromagnetic compatibility – Generic emission standard Part 1: Residential, commercial and light industry.
EN 50 082-1	Electromagnetic compatibility – Generic immunity standard Part 1: Residential, commercial and light industry.

With reference to regulations in the following directives: 73/23/EEC, 89/336/EEC

ABOUT THIS MANUAL

This manual contains a complete list of procedures describing how to install, maintain and using the Culture Myograph System – Model 204CM.

Chapter 1 provides a comprehensive view of the construction and basic features of the complete Culture Myograph System.

Chapter 2 describes step-by-step how to set-up a complete 204CM Culture Myograph System, including all various accessories.

Chapter 3 describes Control of Temperature and Light Intensity

Chapter 4 is a complete manual to the Culture Myograph System. The chapter describes in detail how to use the DMT microscope, how to use and adjust the culture myograph chamber and finally instructions for the daily maintenance of the Culture Myograph System.

Appendixes contain additional information about fuse replacement and system specifications.

UNPACKING THE CULTURE MYOGRAPH SYSTEM

Please take a few minutes to carefully inspect your new Culture Myograph System for any damage, which may have occurred during handling and shipping. If you suspect any kind of damage, please contact DMT immediately and the matter will be pursued as soon as possible. If the packing material appears damaged, please retain it until a possible claim has been settled.

We recommend that you store the packing material for any possible future transport of the Culture Myograph System. In case of transport and the original packing material is unavailable, please contact DMT Sales Department for advice and packing instruction.

After unpacking your new Culture Myograph System, please use the following list to check that the system is complete:

1. Culture Myograph Unit:
   - Culture Myograph Chamber with Chamber Cover
   - 2 Glass Cannulas (Tip outer diameter 125µm)
   - 2 Schott Duran Bottles 25ml

2. DMT Microscope:
   - Temperature Probe
   - Olympus Objective Micrometer including Microscope Objective Holder

3. Culture Myograph Heat Controller Unit:
   - USB-cable for connection to PC
   - Power cord (The shape of the AC plug varies by country; be sure that the plug has the right shape for your location)

4. Pressure Regulator

5. Accessories:
   - Small screwdriver
   - 3 m nylon thread
   - 2 blind plugs including six O-rings (1.07×1.27mm) for the Culture Myograph Chamber
   - 10 O-rings (5.0×1.0mm) for fixation of left glass cannula
   - 10 O-rings (18.0×1.27mm) for cover

6. Software & Manuals:
   - 1 CD with user manual for “Culture Myograph System – Model 204CM”
   - 1 CD with data acquisition software - MyoVIEW II

7. Peristaltic Pump (Optional):
   - Alita Watson Marlow 400, VS2-10R-Midi, 2.5-50rpm

8. Computer (Optional)

CHAPTER 1 - SYSTEM OVERVIEW

1.1 Culture Myograph Unit

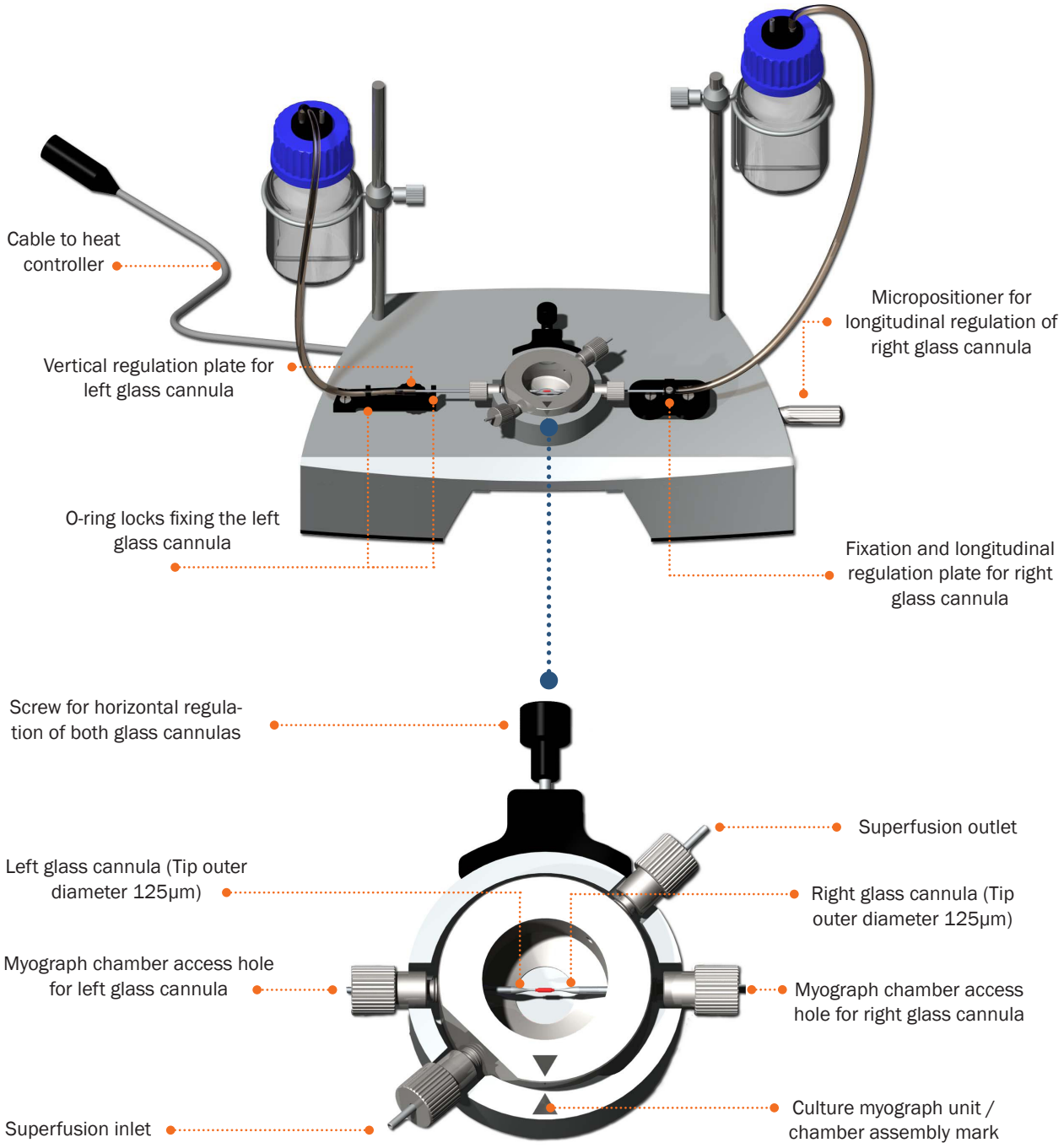

Figure 1.1 *Culture Myograph Chamber Unit*

1.2 DMT Microscope

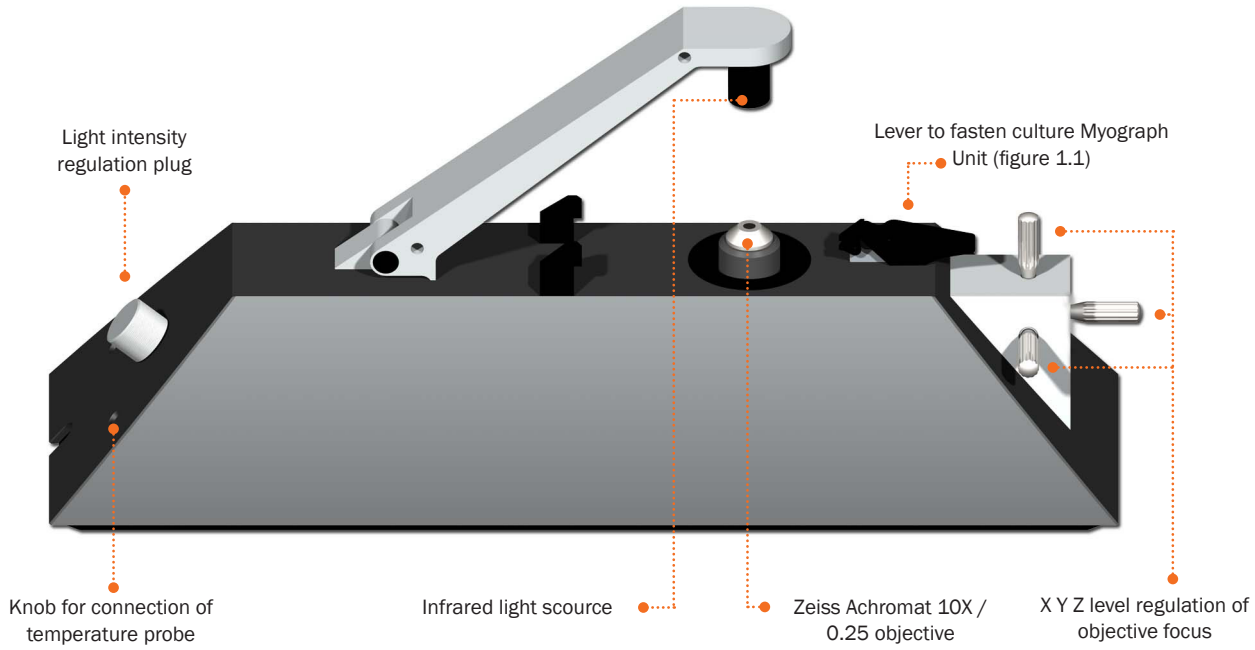

Figure 1.2 *DMT Microscope*

1.3 Culture Myograph Heat Controller

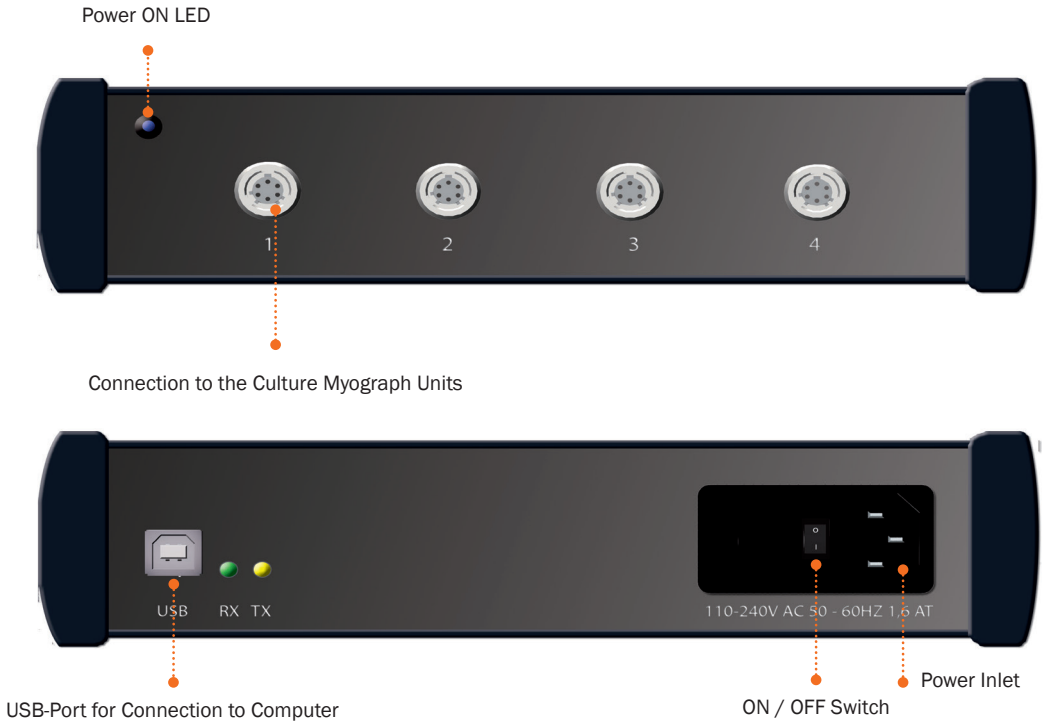

Figure 1.3 *Heat Controller*

CHAPTER 2 - SETTING UP

2.1 The Complete Culture Myograph System - 204CM

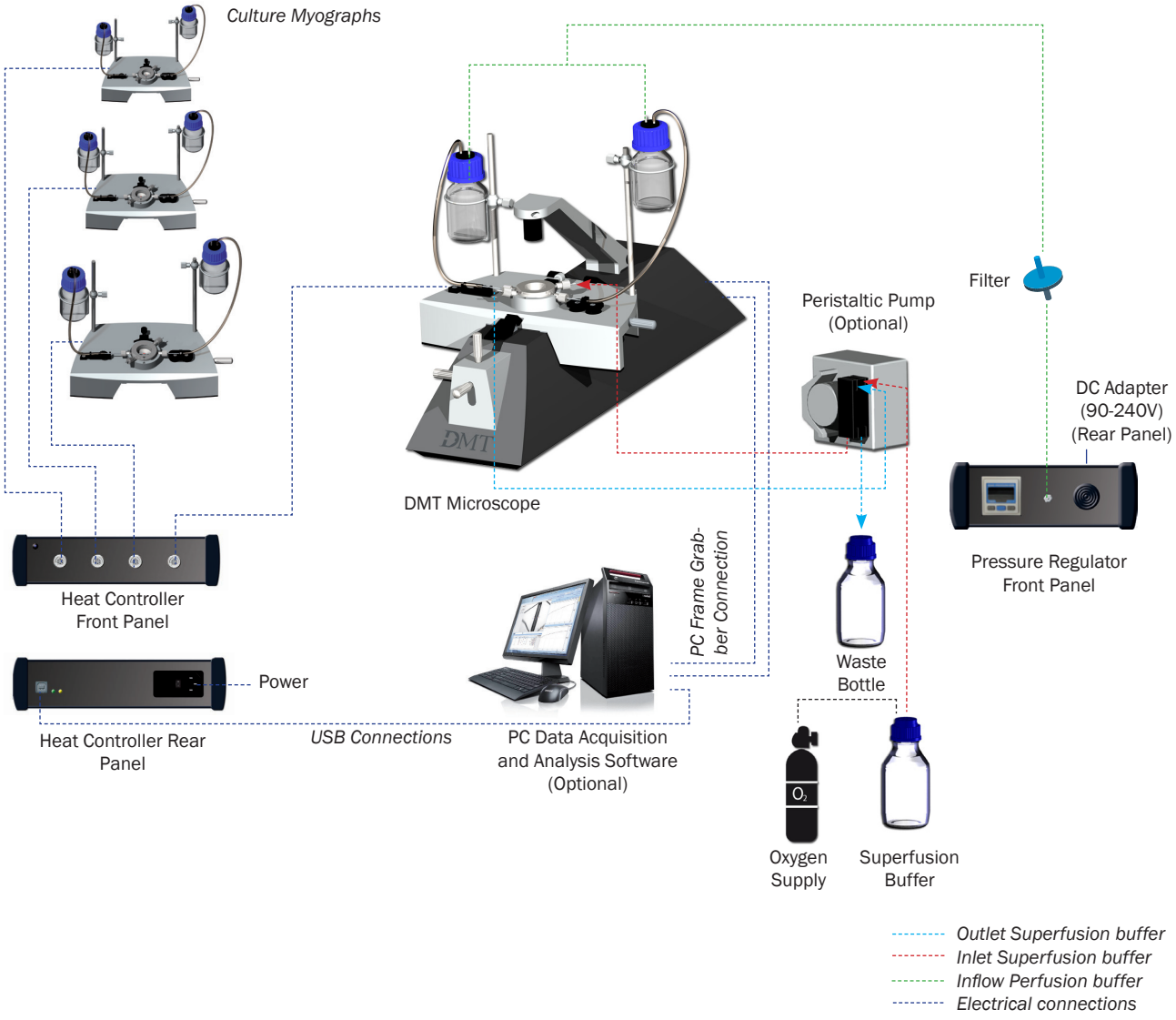

Figure 2.1 *Setting up step by step*

NOTE:
IF YOU HAVE PURCHASED A COMPUTER FROM DMT ALONG WITH YOUR CULTURE MYOGRAH 204CM SYSTEM THE MYOVIEW II HAS ALREADY BEEN INSTALLED ALONG WITH DRIVERS. FOLLOW THE PROCEDURES IN "2.2 SETTING UP THE COMPLETE CULTURE MYOGRAPH SYSTEM 204CM" ON PAGE 13 AND "2.3 INSTALLATION OF MYOVIEW II" ON PAGE 13 TO SET-UP THE CULTURE MYOGRAPH SYSTEM.

IF YOU HAVE NOT PURCHASED A COMPUTER FROM DMT, PLEASE FOLLOW THE PROCEDURES IN "2.3 INSTALLATION OF MYOVIEW II" ON PAGE 13 TO INSTALL MYOVIES II ON YOUR OWN COMPUTER.

2.2 Setting up the Complete Culture Myograph System - 204CM

This section describes how to connect the cables in the culture myograph system as illustrated in figure 2.1 on previous page. Before connecting any of the myograph equipment, ensure that the frame grabber card is installed into the PCI slot of the computer. Please see installation instructions in “Quick Installation Guide for MyoVIEW II”.

1. Ensure that the heat controller is switched off, on the rear panel before proceeding with the connection procedure.
2. Connect the loose cables from the four culture myograph units to the front panel of the heat controller.
3. Connect the power cord to the power inlet on the back panel of the heat controller.
4. Connect the Heat Controller to the PC with the loose USB cable using the USB port on the Heat Controller and a free USB port on the PC.
5. Connect the DMT Microscope to a USB port on the computer using the fixed USB cord.
6. Turn on the power switch of the heat controller.
7. The Power ON LED on the front panel of the heat controller should be lit indicating that the heat controller is on.
8. Connect the DSUB 9-pol cable from the DMT microscope to the 9-pol port on the frame grabber card installed in the computer.
9. Turn on the computer.

2.3 Installation of MyoVIEW II

Please see “Quick Installation Guide for MyoVIEW II”. This guide describes how to install the MyoVIEW II on your computer along with drivers for the digital USB camera.

NOTE:
IF YOU HAVE PURCHASED A COMPUTER FROM DMT ALONG WITH YOUR CULTURE MYOGRAPH 204CM SYSTEM THE DRIVERS AND MYOVIEW II HAVE ALREADY BEEN INSTALLED FOR YOU.

2.4 Experiment Setup

2.4.1 Connecting the Flow-Reservoirs

To connect the flow reservoirs with the glass cannulas and the Pressure Regulator, DMT recommends PharMed™ tubes (Product No. 100127):

NOTE:
A STERILE MICRO FILTER (0.20µm) IS CONNECTED BETWEEN THE PRESSURE MANOMETER RESERVOIR AND THE TWO PERFUSION RESERVOIRS TO PREVENT CONTAMINATION OF THE PERFUSION BUFFER.

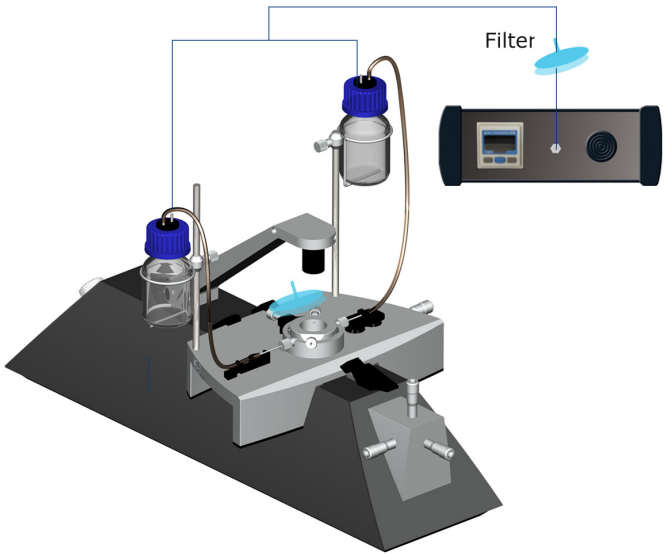

Figure 2.2 *Illustration of how to connect flow reservoirs.*

2.4.2 Perfusion Flow Control

Regulating the difference in height between the two flow reservoirs allows control of the perfusion flow velocity. The principle is illustrated in figure 2.3 below. In figure 2.3 A below the two reservoirs are equal in height and no flow will occur. In figure 2.3 B below the difference in height reveals a flow from the right reservoir to the left reservoir.

NOTE:
TO ENABLE THE PERFUSION FLOW, IT IS IMPORTANT THAT THE RIGHT GLASS CANNULA IS CONNECTED TO THE LONG STEEL PIPE IN THE RIGHT RESERVOIR (MARKED BY THE ARROW IN FIGURE 2.3 A BELOW).

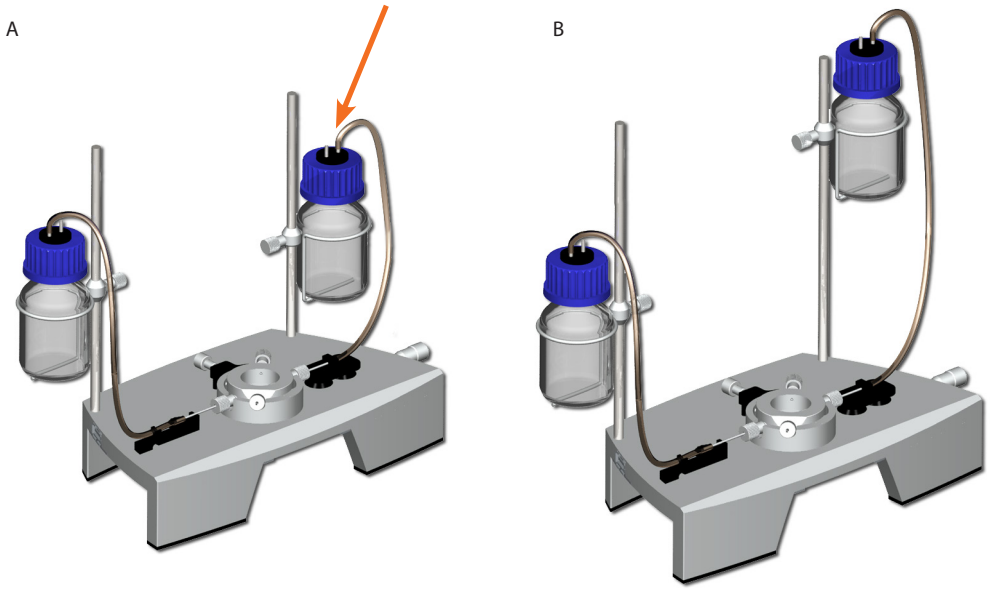

Figure 2.3 A and B *Illustration of perfusion flow control*

2.4.3 Setup and Control of Superfusion Flow

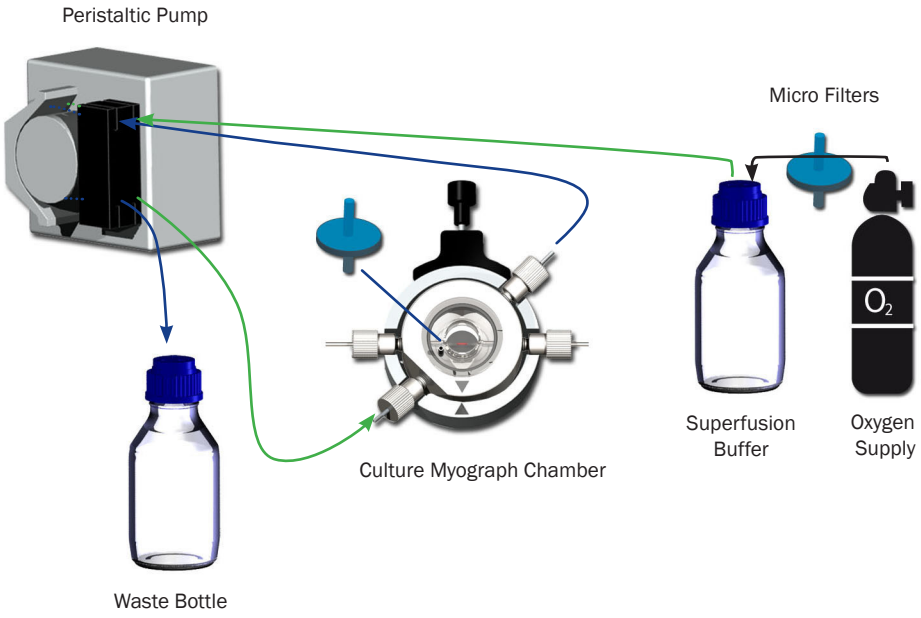

Figure 2.4 *Illustration of perfusion flow control*

The superfusion circuit consists of an inflow and an outflow from the culture myograph chamber. Both flows are driven by the same peristaltic pump, which leads the superfusion buffer from the reservoir to the culture myograph chamber and finally to a waste bottle. The small steel pipe on the culture myograph chamber cover is connected to a sterile micro filter (0.20µm). The small pipe works as a breathing valve to prevent over pressure in the culture myograph chamber.

The superfusion buffer reservoir is continuously aerated with a mixture of 95% O_2 and 5% CO_2. The gas passes through a sterile micro filter (0.20µm) to prevent contamination of the sterile superfusion buffer.

CHAPTER 3 - DMT 204CM CONTROL PROGRAM

3.1 Control of Temperature and Light Intensity

In addition to MyoVIEW II, a program is installed on the computer allowing temperature and light intensity control of the Culture Myograph 204CM system.

To run the DMT 204CM control program, please select the “204CM controller” which was copied to your desktop during installation. “204CM controller” can also be found in START->All Programs->DMT->204CM

NOTE:
MAKE SURE THAT THE HEAT CONTROLLER IS TURNED ON AND THE MICROSCOPE IS CONNECTED TO THE FRAME GRABBER, BEFORE RUNNING THE 204CM CONTROLLER.

3.1.1 Connecting the Heat Controller and the Microscope

When the 204CM Controller is started it will automatically connect to the Microscope and Heat Controller. In the About Tab, the status of the COM connections is shown. If the connection fails please check that the Microscope and Heat Controller is turned on, and that the USB cables is connected.

CHAPTER 4 - CULTURE MYOGRAPH MANUAL

4.1 The Culture Myograph Unit

The culture myograph unit is placed on the DMT microscope as illustrated in figure 4.1 below. Press the black lever to lock the culture myograph unit onto the DMT microscope.

4.1.1 Glass Cannula Adjustment

The glass cannulas are adjustable in all X Y Z directions. The adjustments are illustrated in figure 4.1 below.

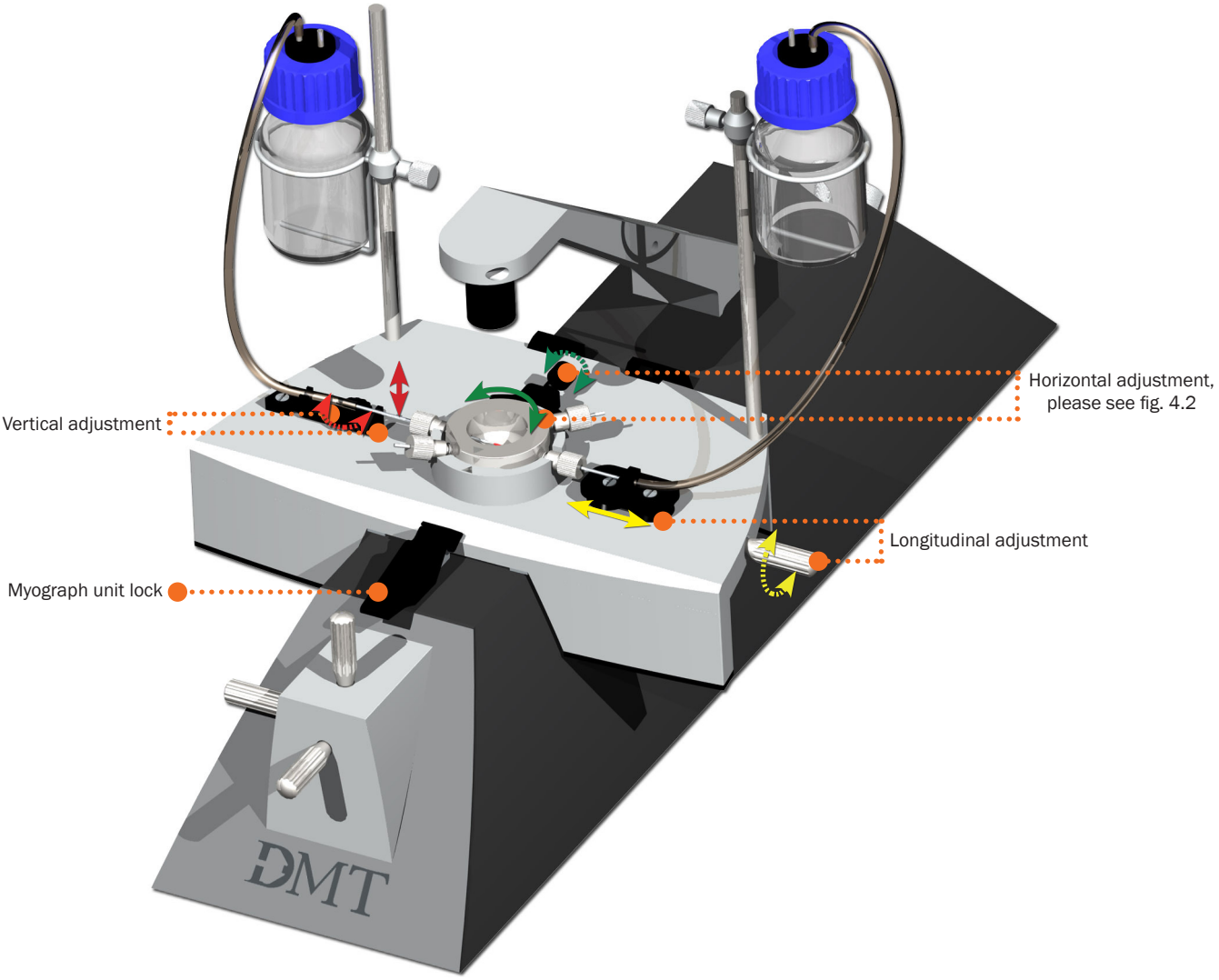

Figure 4.1 *Illustration of glass cannula adjustment*

Horizontal Adjustment:
Horizontal adjustment is performed as illustrated in figure 4.1 on previous page and figure 4.2 below.

Gently loosen the screw fixing the chamber and carefully move the myograph chamber in either clockwise or counter clockwise direction to adjust the horizontal alignment of the glass cannulas.

Horizontal Adjustment Screw:
Loosen this screw and carefully turn clockwise or counter clockwise. Observe how the pippettes move in horisontal plane.

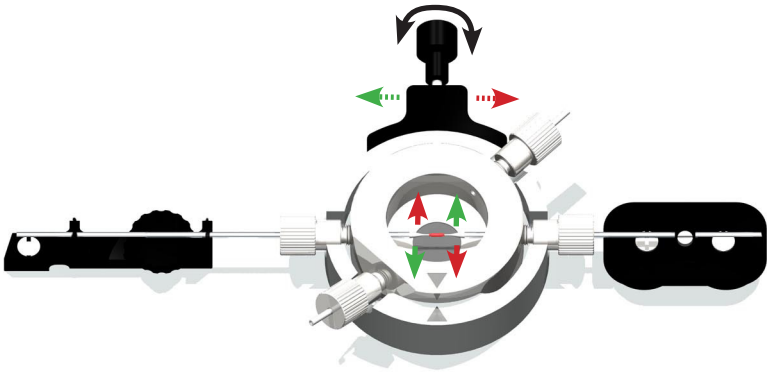

Figure 4.2 *Horizontal adjustment of glass pipettes*

Vertical Adjustment:
Vertical adjustment is performed as illustrated in figure 4.1 on previous page.

Gently turn the black dish, underneath the left glass cannula, clockwise or counter clockwise to adjust the vertical alignment of the glass cannulas.

Longitudinal Adjustment:
Longitudinal adjustment of the glass cannulas is performed using the micro positioner on the right side of the culture myograph unit.

4.1.2 Removal and Mounting of Glass Cannulas and Chamber

Replacement of glass cannulas and demounting of the culture myograph chamber for sterilisation are common routines in culture myograph experiments. The procedure is illustrated in figure 4.3 below.

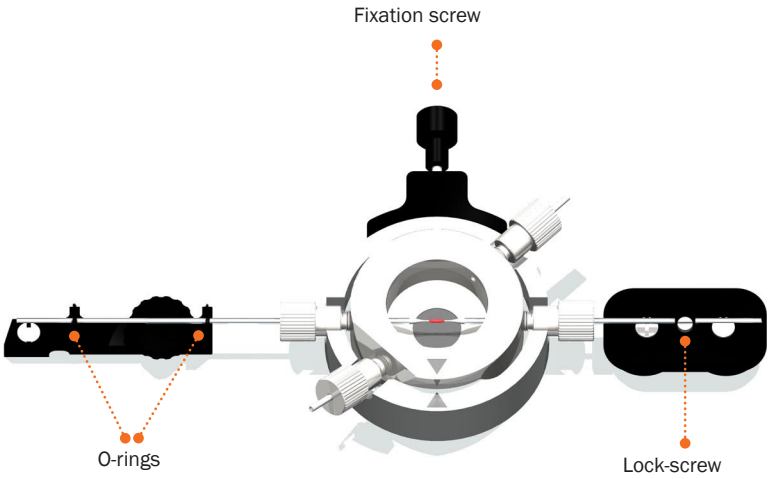

Figure 4.3 *How to free the glass cannulas from the culture myograph unit*

Removal of Left Glass Cannula:
Carefully remove the two O-rings (marked by the two arrows) to free the glass cannula from the culture myograph unit. Then carefully unscrew the locknut and remove both locknut and glass cannula.

Removal of right Glass Cannula:
Gently remove the little screw (marked by the arrow) to free the glass cannula from the culture myograph unit. Then carefully unscrew the locknut and remove both locknut and glass cannula.

Removal of Culture Myograph Chamber:
After removal of both glass cannulas, loosen the horizontal adjustment screw to free and remove the chamber from the culture myograph chamber.

Mounting of all parts is performed the same way, but in opposite direction.

4.1.3 Changing O-rings

Each locknut on the culture myograph chamber is fitted with rubber O-rings (ø1.07 x 1.27mm) to keep the chamber tight. These O-rings will from time to time need to be replaced. The procedure is illustrated in figure 4.4 below.

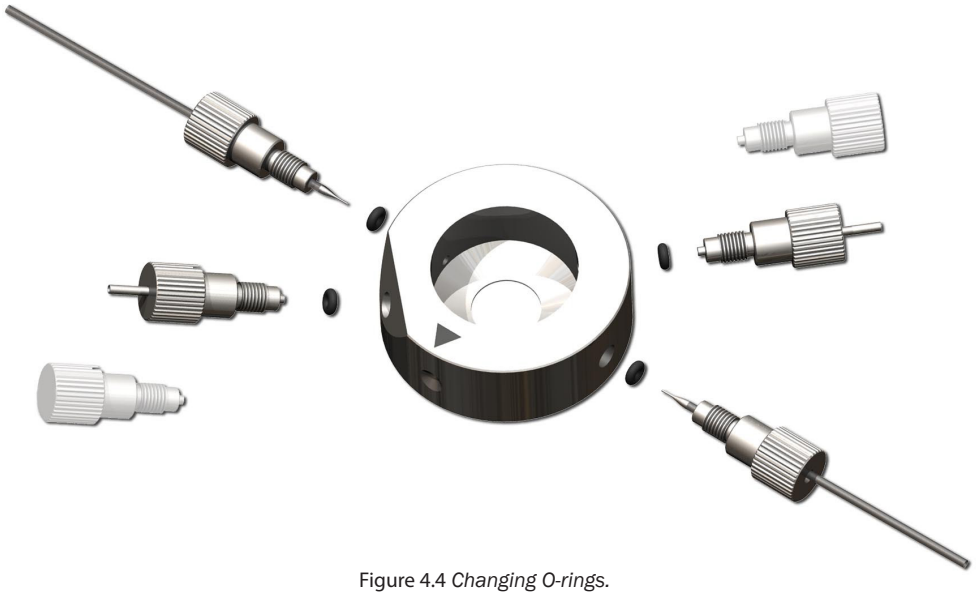

Figure 4.4 *Changing O-rings.*

- Unscrew the locknut to reveal the O-ring and remove it carefully using small forceps or a similar tool.
- Insert the new O-ring into the chamber hole and push it back until it makes contact with the bottom of the hole.
- Carefully insert the glass cannula through the locknut and place it in the chamber hole. Gently tighten the locknut.

NOTE:
BE CAREFUL NOT TO DAMAGE THE GLASS CANNULA TIPS WHEN DISASSEMBLING THE CULTURE MYOGRAPH CHAMBER.

4.2 The DMT Microscope

The DMT microscope is an invert microscope equipped with a Zeiss Achromat 10X / 0.25 objective and build-in digital USB CCD camera. The objective is adjustable in all X Y Z directions using the three micro positioners on the front as illustrated in figure 4.5 below.

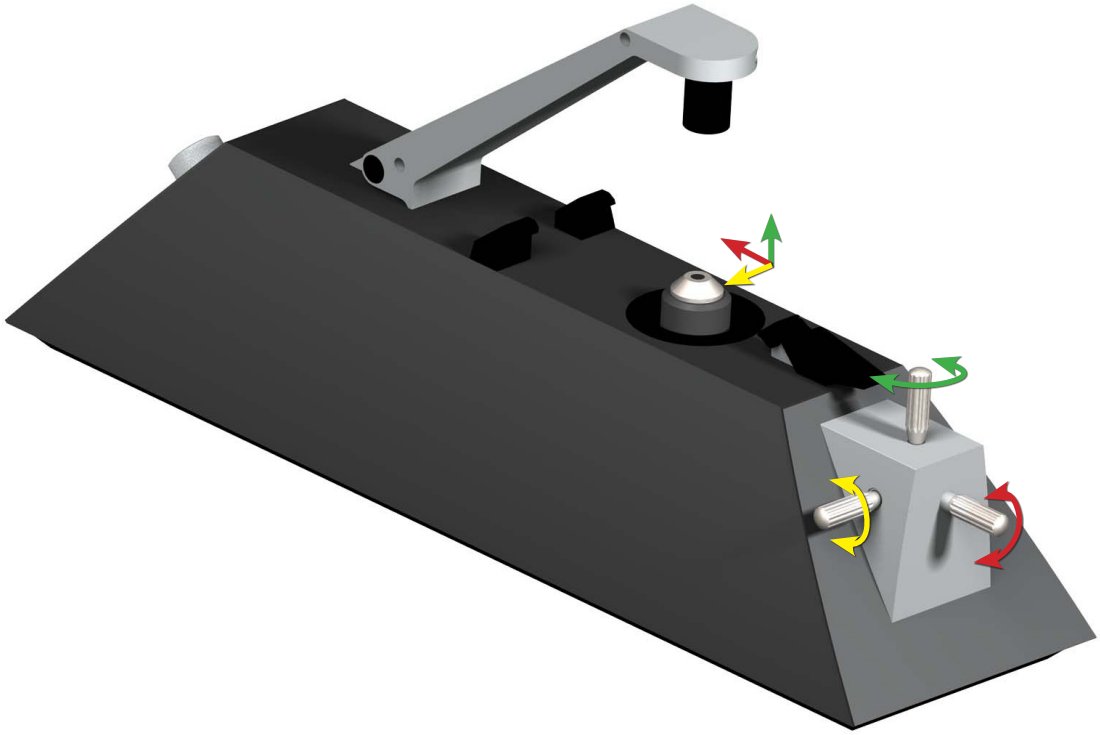

Figure 4.5 *How to adjust the position and focus of the microscope in the X Y Z directions.*

MyoVIEW II obtains an image by measuring the differences in light intensity passing through the walls of a vessel segment mounted in the culture chamber. Traditionally a white light source is sufficient for such a purpose, but has one major disadvantage. A white light source makes the data acquisition and analysis very sensitive to changes in white light intensity from the surroundings, such as ambient light and sunlight. To avoid the surroundings influencing the data acquisition and analysis, the DMT microscope has a built-in infrared light source.

4.3 Pressure Regulator

The Pressure Regulator is connected to the culture myograph system to generate a pressure on the mounted vessel from the perfusion buffer. The pressure is adjusted using the knob and air release valve shown in figure 4.6 below.

Figure 4.6 *Pressure Regulator*

4.4 Culture Myograph Maintenance

The Culture Myograph System – Model 204CM is a very delicate and sophisticated piece of research equipment. In order to keep it working at its best, DMT recommend that the following sections are read carefully and that the instructions are followed at all times.

DMT STRONGLY RECOMMENDS THAT THE MYOGRAPH CHAMBER AND SURROUNDINGS BE CLEANED AFTER EACH EXPERIMENT.

After a "normal" experiment use the following procedure to clean the myograph chamber and glass cannulas:

1. Fill up the myograph chamber to the edge with an 8% acetic acid solution and allow it to stand for a few minutes to dissolve calcium deposits and other salt build-up. Use a swab stick to mechanically clean all chamber surfaces.

2. Remove the acetic acid and wash the myograph chamber and glass cannulas several times with double distilled water.

3. It is hard to remove any kind of hydrophobic reagent used by using step 1. and 2., try incubating the chamber and glass cannulas with 96% ethanol or a weak detergent solution (e.g. Treepol).

4. To remove more resistant or toxic chemicals, incubate the myograph chamber and glass cannulas with 1M HCl for up to 1 hour. In exceptional cases incubate the chamber and supports with a up to 3M HNO_3 solution for about 15 minutes.

5. Wash the myograph chamber and glass cannulas several times with double distilled water.

IMPORTANT NOTES:

TO STERILIZE THE CULTURE MYOGRAPH CHAMBER AND GLASS CANNULAS, USE A STANDARD AUTOCLAVE PROCEDURE.

BE VERY CAREFUL USING STEP 3 AND 4 REPEATEDLY AS STRONG REAGENTS CAN CAUSE EXTREME DAMAGE TO THE MYOGRAPH UNIT.

BE VERY CAREFUL NOT TO DAMAGE THE GLASS CANNULAS DURING THE CLEANING PROCEDURE.

In cases of red or brown discolorations appearing on the chamber sides, the following cleaning procedure will work in most cases:

1. Incubate the myograph chamber for 30 minutes with 20µl of a 2mM T-1210 Tetrakis-(2-pyridylmethyl)-ethylenediamine solution dissolved in double distilled water.

2. Use a swab-stick to mechanically clean all the affected surfaces during the last 15 minutes of the incubation period.

3. Wash the myograph chamber several times with double distilled water.

4. Incubate the myograph chamber with 96% ethanol for 10 minutes while continuing the mechanical cleaning with a swab-stick.

5. Remove the ethanol solution and wash a few times with double distilled water. Incubate the myograph chamber with an 8% acetic acid solution for 10 minutes and continue the mechanical cleaning with a swab-stick.

6. Wash the myograph chamber several times with double distilled water.

CHAPTER 5 - GETTING STARTED

5.1 Dissection Protocol for Small Mesenteric Arteries

The culture myograph technique is versatile in that a large variety of physiological and pharmacological studies of ring preparations from different species can be performed. Mostly, the culture myograph is used for investigation of small blood vessels and as an example this chapter describes the dissection of rat mesenteric arteries.

1. A laboratory rat is euthanized in accordance to the local national law and regulations. A midline laparotomy is performed to expose the mesenteric bed.

2. Use scissors to remove about 10cm of intestine along with its feeding vasculature, including part of the superior mesenteric artery. Be careful not to damage the vasculature during this procedure. The proximal end of the intestine section must be about 10cm from pylorus. Make a cut in the proximal end of the intestine for later identification.

3. Place the excised intestine section in a Petri dish (about 9cm in diameter) coated with a 5mm thick layer of Sylgard at the bottom to hold the fixing pins. Immediately fill the Petri dish with cold PSS well prebubbled with carbogen (see Chapter 5.3). The dissection is performed without further oxygenation of the PSS.

4. Pin down the proximal end of the intestine section on the left-hand side of the Petri dish without stretching the vessels. Pin down the remaining of the intestine section in an anti-clockwise direction. In this configuration (proximal end at the left side, distal end at the right side and running anti-clockwise from proximal to distal side) the feeding vasculature is on the far side of the intestine and the veins are usually uppermost.

5. Select the vessel segment to be investigated (Fig. 5.1). First time myograph users are recommended to start dissecting and mounting vessel segments from the first or second branch from the superior mesenteric artery (approximate internal diameter 200-300µm).

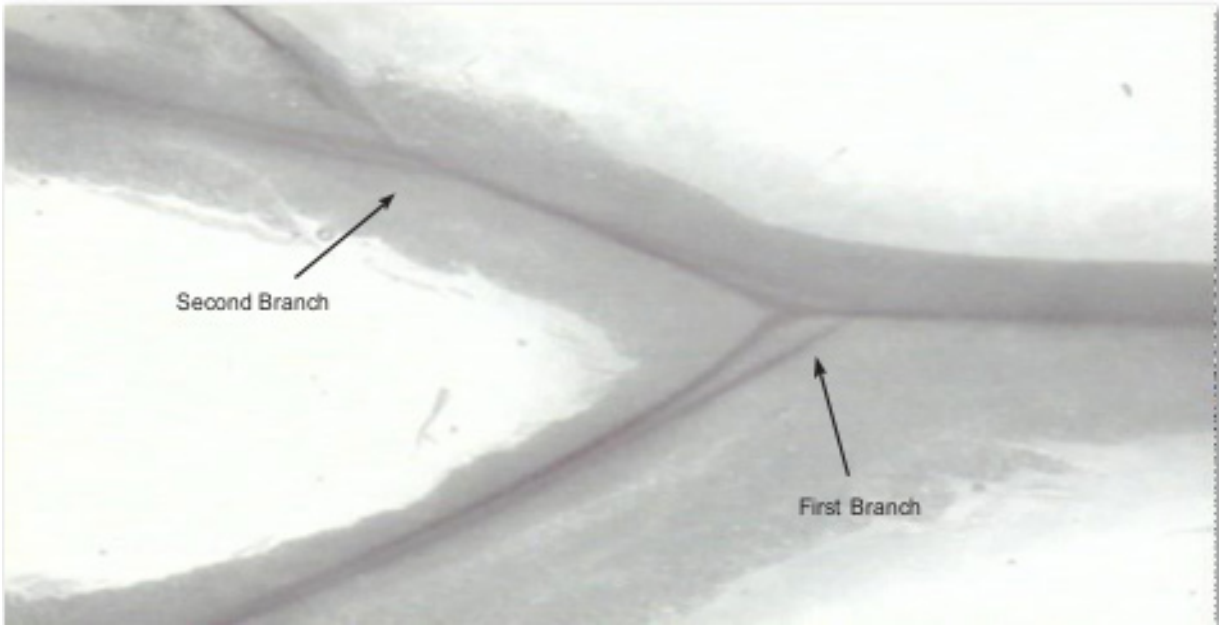

Figure 5.1 Branch of the mesenteric arteries

6. Use high quality forceps and ocular dissection scissors to dissect the vessel segment of interest. Start cutting through the mesenteric membrane along both sides of the vessel, about 1-2mm from the vessel. To avoid accidentally cutting the artery always cut along the length of the vessels and never perpendicular to them (Fig. 5.2 A-B).

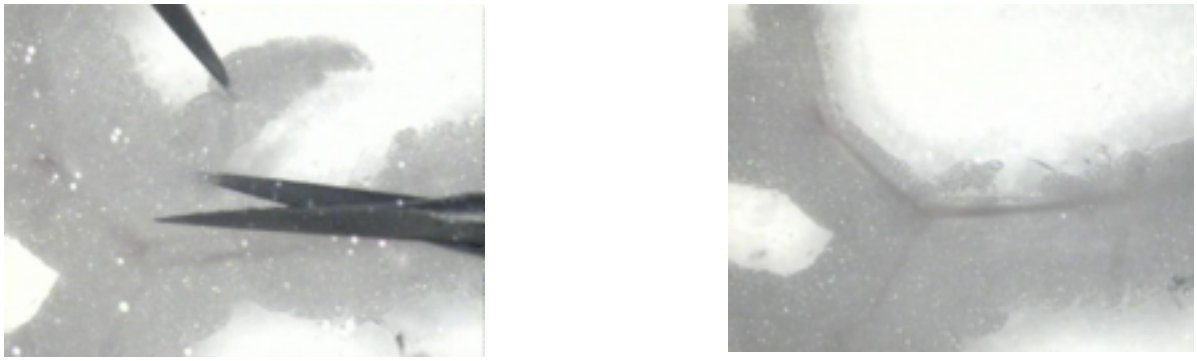

Figure 7.2 Removal of adipose tissue around the area of interest

7. Dissect away as much adipose tissue as needed around the vessels to distinguish between the artery and vein. The artery can easily be identified by the following characteristics (Fig 5.3):

- The branch points of arteries are V-shaped whereas those of veins are more U-shaped.
- The arterial wall contains a thick layer of smooth muscle cells compared to the vein wall, which only contains a single or a few layers of smooth muscle cells. The histological difference is clearly visible in the stereo microscope.
- If you still have difficulty and the vein and artery still contain some blood then try to move the blood forward by very gently squeezing the vessels with a forceps. In the artery the blood will run back quickly whereas in the vein the blood will run back very slowly if it even does so. Note, it is important that you perform this on vessels other than those you will use as this procedure damages the vessels.

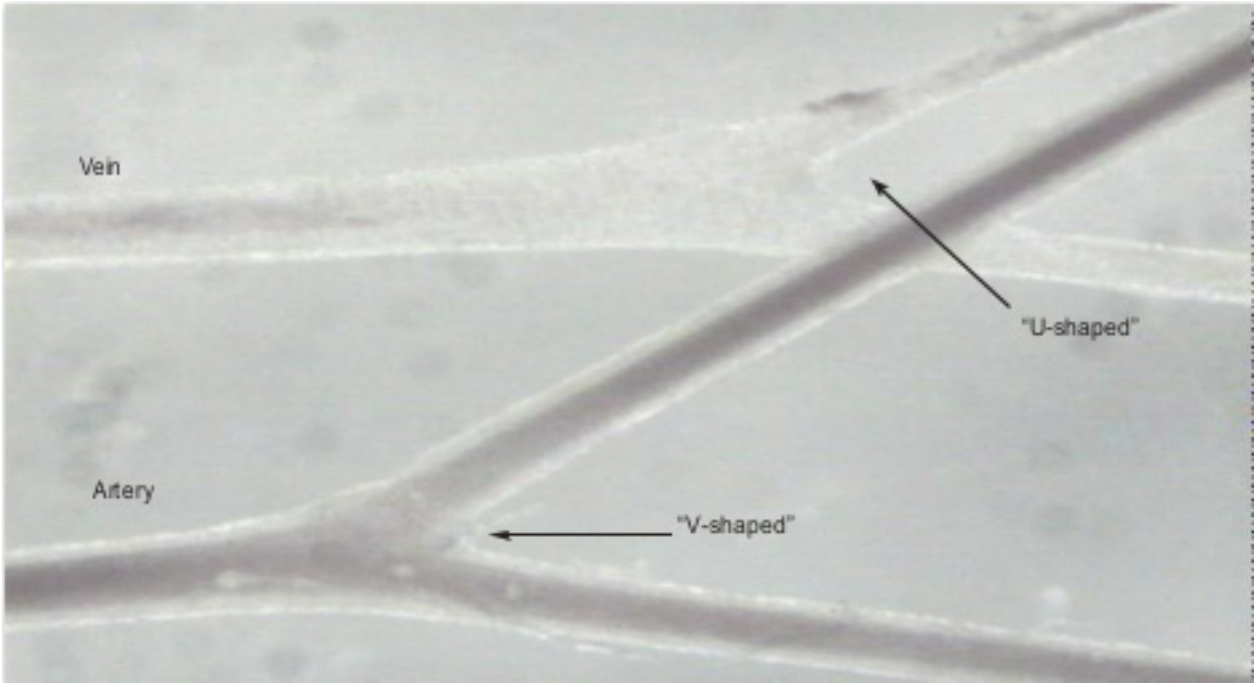

Figure 5.3 Distinguishing between artery and vein

8. Dissect away the vein using scissors to cut the adipose and connective tissue between the artery and vein. One method is to cut the vein in one position and afterwards gently to pull the vein away from the artery. In this way, a fine membrane of connective tissue becomes visible between the adipose tissue and the artery. Carefully cut the fine membrane to remove the vein and adipose tissue while avoiding any direct contact between the scissor and artery (Fig. 5.4 A-B).

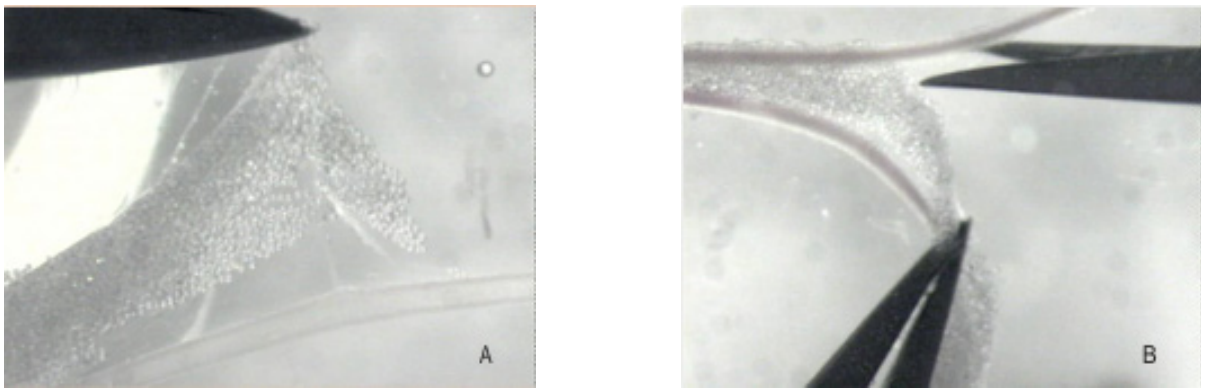

Figure 5.4 Removal of vein

9. Clean the artery by removing any remaining adipose or connective tissue. Gently pull away adipose or connective tissue to make the connective tissue membrane become visible. Cut the membrane to remove the tissue.

10. Cut the distal end of the artery section to be investigated. Afterwards cut the proximal end while ensuring that the vessel segment has the correct length (Fig. 5.5 A-C).

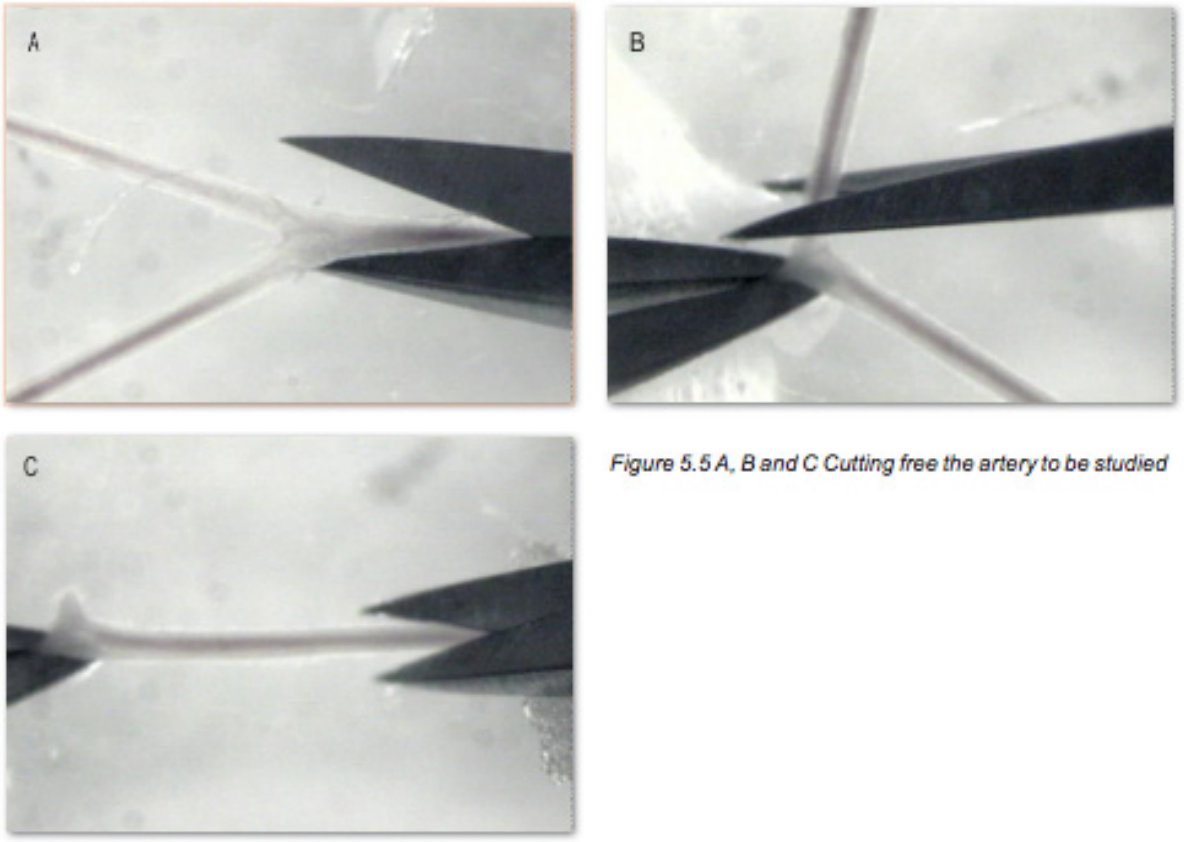

Figure 5.5 A, B and C Cutting free the artery to be studied

5.2 Mounting Protocol for Small Arteries

This section shortly describes the basic technique of mounting small vessels in the culture myograph:

- One end of the vessel is care-fully mounted and secured (with two fine nylon sutures) to one of the hollow glass micro- cannulas.
- The lumen is flushed gently with PSS to remove any blood or debris.
- The second end of the vessel is then mounted and secured to the second micro-cannula.
- The PSS in the myograph chamber is gradually warmed to 370C over a period of 15 minutes.

Figure 5.6 Mounting of vessel in the culture myograph

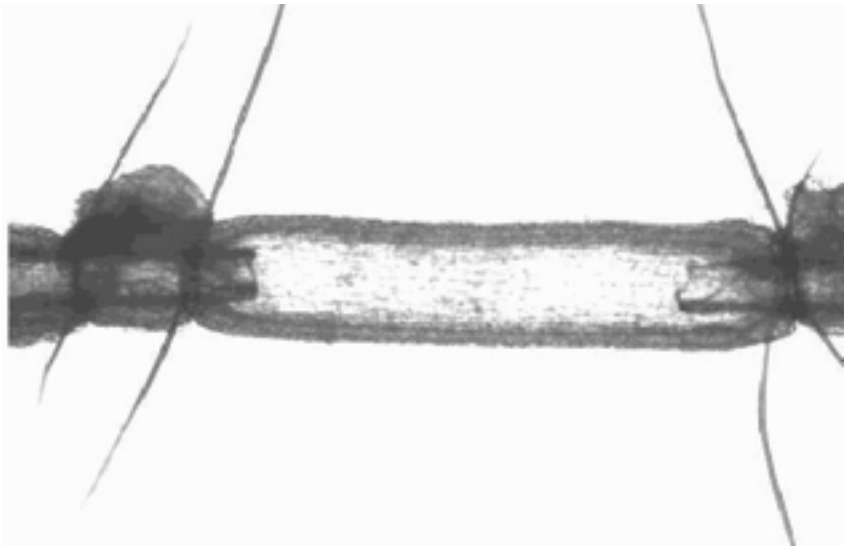

Figure 5.7 Vessel secured to the two glass

5.3 Buffer Recipes

Physiological Saline Solution (PSS)

1x PSS:

Chemical	Mol.Wt	mM	g/0.5L	g/L	g/2L	g/4L
NaCl	(58.45)	130	3.799	7.598	15.20	30.39
KCl	(74.557)	4.7	0.175	0.35	0.70	1.40
KH_2PO_4	(136.09)	1.18	0.08	0.16	0.32	0.64
$MgSO_4 \cdot 7H_2O$	(246.498)	1.17	0.145	0.29	0.58	1.16
$NaHCO_3$	(84.01)	14.9	0.625	1.25	2.50	5.00
Glucose	(180.16)	5.5	0.5	1.00	2.00	4.00
EDTA	(380)	0.026	0.005	0.01	0.02	0.04
$CaCl_2$	(110.99)	1.16	0.8ml	1.6ml	3.2ml	6.4ml

(1.0 M solution)

1. Make a 1.0M solution of CaCl2 (110.99) in double-distilled H O. Filter-sterilize the calcium solution through a 0.22 µm filter. The sterilized solution can be stored in the refrigerator for up to 3 months.

2. Dissolve all the chemicals except the CaCl2 in approximately 80% of the desired final volume of double distilled H2O while being constantly stirred. For example, if 1 litre of PSS is to be made, then dissolve all the chemicals in 800ml of double distilled H2O.

3. Add the appropriate volume of 1.0M CaCl2 for the total volume of PSS being made (for example, 1.6ml of 1.0M CaCl2 for 1 litre of buffer). Continue to stir the PSS while the CaCl2 is being added.

4. Bring the solution up to the final volume with double-distilled H2O. Continue to stir the solution until the EDTA is fully dissolved. This takes about 15 minutes at room temperature.

5. Aerate the solution with carbogen for about 20 minutes.

2!

Chemical	Mol.Wt	mM	g/0.5L	g/L	g/2L	g/4L
NaCl	(58.45)	130	3.799	7.598	15.20	30.39
KCl	(74.557)	4.7	0.175	0.35	0.70	1.40
KH_2PO_4	(136.09)	1.18	0.08	0.16	0.32	0.64
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4. Bring the solution up to the final volume with double-distilled H2 O. Continue to stir the solution until the EDTA is fully dissolved. This takes about 15 minutes at room temperature.

Before use:

5. Dilute the 25 x PSS stock solution 1:25 using double distilled H2O

6. Add: 91g/L Glucose 100 g/L NaHCO3

7. Aerate the solution with carbogen (95%O2+5%CO2) for at least 20 minutes. If necessary wait further for the pH of the buffer to reach 7.4 pH.

High potassium Physiological Saline Solution (KPSS)

1x 60mM KPSS:

Chemical	Mol.Wt	mM	g/0.5L	g/L	g/2L	g/4L	
NaCl (58.45)	74.7	2.18	4.37	8.73	17.46		
KCl (74.557)	60	2.24	4.47	8.95	17.89		
KH2PO4	(136.09)	1.18	0.08	0.16	0.32	0.64	
MgSO4 7H2O	(246.498)		1.17	0.145	0.29	0.58	1.16
NaHCO3	(84.01)	14.9	0.625	1.00	2.00	5.00	
Glucose	(180.16)	5.5	0.5	1.00	2.00	4.00	
EDTA (380)	0.026	0.005	0.01	0.02	0.04		
CaCl2 (110.99)	1.6	0.8ml	1.6ml	3.2ml	6.4ml		

(1.0 M solution)

1. Make a 1.0M solution of CaCl2 (110.99) in double-distilled H2O. Filter-sterilize the calcium solution through a 0.22 µm filter. The sterilized solution can be stored in the refrigerator for up to 3 months.

2. Dissolve all the chemicals except the CaCl2 in approximately 80% of the desired final volume of double distilled H2O while being constantly stirred. For example, if 1 litre of PSS is to be made, then dissolve all the chemicals in 800ml of double distilled H2 O.

3. Add the appropriate volume of 1.0M CaCl2 for the total volume of PSS being made (for example, 1.6ml of 1.0M CaCl2 for 1 litre of buffer). Continue to stir the PSS while the CaCl2 is being added.

4. Bring the solution up to the final volume with double-distilled H2O. Continue to stir the solution until the EDTA is fully dissolved. This takes about 15 minutes at room temperature.

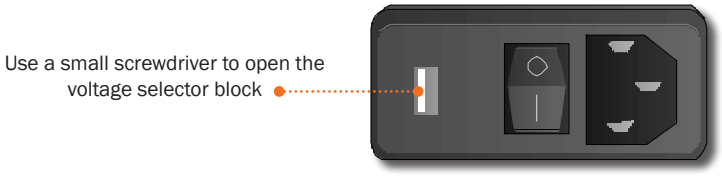
5. Aerate the solution with carbogen (95% O2 + 5% CO2) for about 20 minutes.

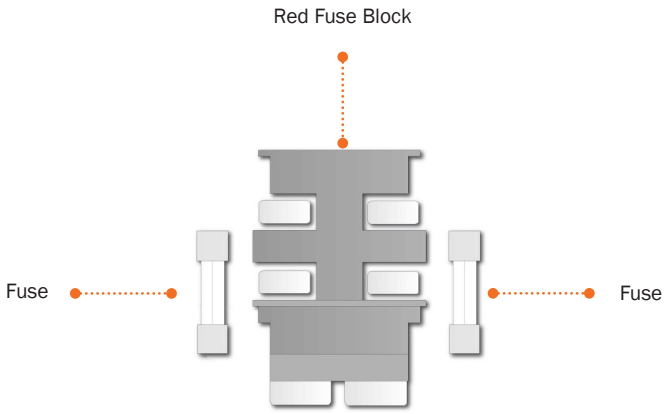
APPENDIX 1 - FUSE REPLACEMENT

The main fuse of the myograph system is placed inside the power inlet on the Heat controller. If the fuse blows it is easily changed using the following procedure.

IMPORTANT:
IF THE FUSE NEEDS TO BE CHANGED MAKE SURE THAT THE REPLACEMENT FUSE IS EQUAL TO THE ONE BLOWN.

Specifications: T2A / 250V, 6.3 x 32mm





APPENDIX 2 - SYSTEM SPECIFICATIONS

Technical specifications

Chamber Unit	
Vessel size:	>60 µm
Chambers:	Four
Chamber volume:	Max. 2 ml
Cannula ports:	O.D. 1.2 mm
Superfusion ports:	Built-in
Base window:	10 mm diameter type II coverslip
Chamber material:	Acid-resistant stainless steel
Chamber cover:	Removable with gasket and access port
Chamber Stage	
Alignment:	Two cannula holders
Adjustment:	X, Y & Z
Flow:	25ml bottles for flow adjustment
Temp. range:	Ambient temp - 50 °C
Temp. control:	Via PC software
Temp. probe:	Included
Microscope Stand	
Camera:	USB ½"
Objective:	10x
Adjustment:	X, Y & Z directions
Light source:	Infrared LED
Voltage:	100 to 240 VAC (auto) 50/60 Hz
Pressure Regulator	
Pressure range:	0 - 300 mmHg
Pressure source:	Atmospheric pressure
Heat controller	
Temperature range:	Max 50 °C
Pump	
Peristaltic pump:	2.5 - 50 rpm (for superfusion of the chamber) 4-8 channel versions available

Optional accessories

FlowMeter	
- range:	15 µl/min to 4000 µl/min